

RITUXIMAB IN NEUROLOGICAL DISEASE: PRINCIPLES, EVIDENCE AND PRACTICE IN AMARA CITY AT ALSADER TEACHING HOSPITAL IN MISAN

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Abstract

Rituximab is a popular monoclonal antibody that kills B cells. There are no treatment recommendations and it is not approved for use in treating neurological disorders. However, it is an appealing substitute for traditional immunomodulatory drugs as a swiftly acting, targeted therapy with mounting evidence of efficacy and tolerance in numerous neuroinflammatory conditions. The goal of this hands-on review is to clarify the fundamental ideas behind therapeutic monoclonal antibody B-cell depletion. We discuss the scientific support for the use of rituximab in neurological illnesses as well as dose, monitoring, safety, treatment failure, and its application in unique situations such concurrent viral hepatitis, pregnancy, and breastfeeding. We offer a patient information booklet, administration guide, and checklist that can be customized for local use. Lastly, we assess the security.

Introduction

This article discusses some of the fundamental ideas of B-cell depletion with monoclonal antibodies, which are pertinent to neurologists, as well as some of the practical aspects of prescribing rituximab. Skip to the appropriate tables near the conclusion of the article if you're looking for an administration guide for rituximab, a quick summary of the indications and supporting data, anticipated side effects, or specific prescription situations. An illustration of a patient information sheet and an administration checklist are supplied.

The involvement of B cells in neurological illness

B-cells produce proinflammatory and antiinflammatory cytokines, release antibodies, deliver antigen, and control the immune response. Only 2.5% of the total B-cell population, primarily made up of memory and naive mature B-cells, are found in the peripheral circulation; the remaining 95% are found in bone marrow and lymphoid tissue. 1 Any immunoglobulin class (G, M, A, D, or E) or subclass (e.g., IgG1-4) may contain antibodies, and each has a different function. Myasthenia gravis with antibodies against the acetylcholine receptor (AChR) or muscle-specific tyrosine kinase (MuSK) (IgG4), neuromyelitis optica spectrum disorders (NMOSD) with antibodies against the aquaporin-4 water channel (primarily IgG1), and autoimmune encephalitis with antibodies are examples of disorders in which autoantibodies are almost certainly pathogenic. Either the leucine-rich glioma inactivated-1 (LGI1) or N-methyl-D-aspartate receptor (NMDAR),



which are primarily IgG1 receptors (mainly IgG4). As shown by oligoclonal IgG bands in the cerebral fluid, meningeal-based ectopic B-cell follicles next to regions of localized cortical demyelination², and the effectiveness of B-cell-depleting therapy for MS, B-cells also play a significant role in the pathophysiology of multiple sclerosis (MS).

Surface markers on B-cells

The B-cell transmembrane proteins CD19 and CD20. They can serve as surface indicators and pharmacological targets (in flow cytometry to quantify B-cell populations and assess treatment response). During B-cell development, CD19 is produced more frequently than CD20, although neither marker is present on long-lived plasma cells (figure 1). In healthy adults, the percentage of CD19+ or CD20+ B-cells in the total circulating lymphocyte population ranges from 12% to 22% (the absolute reference range is 50–500 cells/mm³).

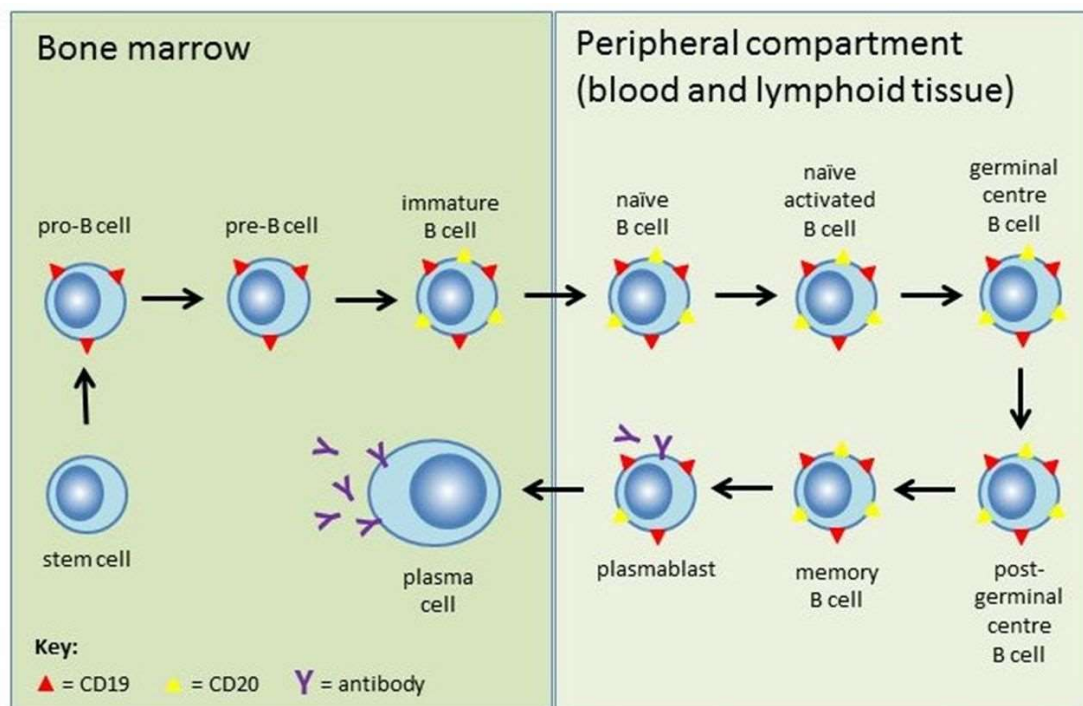


Figure 1

Stages of B-cell development and surface marker expression are shown in Figure 1. In the bone marrow, pluripotent haematopoietic stem cells grow into naïve mature B cells. When they reach the spleen and lymph nodes, secondary lymphoid organs, they travel there where they are activated by antigens in the lymph fluid and develop into memory B-cells or plasmablasts. While plasmablasts develop into antibody-secreting plasma cells that live in the bone marrow or lymphoid tissue, memory B-cells either circulate in the blood or stay in germinal centers. At the plasmablast stage, CD20 (yellow triangles) disappears after first appearing at the stage of immature B-cells. The great majority of plasmablasts and virtually all plasma cells, which are responsible for producing most antibodies, do not express CD20. Wider expression of CD19 (red triangles) is seen from the pro-B-cell stage.

Memory B-cells and some other immune cell types express CD27. Memory B-cells only express

CD19 and CD27 in combination. This long-lived B-cell subgroup, which can quickly differentiate into high-affinity plasma cells after repeated antigen exposure, may be a key target in the therapy of autoimmune neurological diseases. 3 4

Monoclonal antibodies that eliminate B cells

Immunoglobulins called monoclonal antibodies are created by a single clone of hybridoma cells (antigen-specific plasma cells fused with myeloma cells). Their two identical fragment antigen binding (Fab) domains bind to a single epitope, and their fragment crystallisable (Fc) domain stimulates the immune system. A range of neoplastic and autoimmune illnesses can be treated with highly targeted immunotherapy because cells that express that epitope are eliminated. With the exception of mature plasma cells that secrete antibodies, available B-cell-depleting monoclonal antibodies with Fab domains that target CD20 or CD19 selectively eliminate the circulating B-cell population. Those used to treat neuroinflammatory disorders are depicted in figure 2.

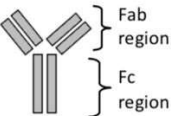
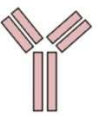
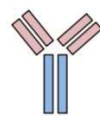
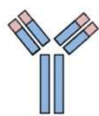
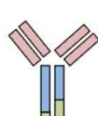
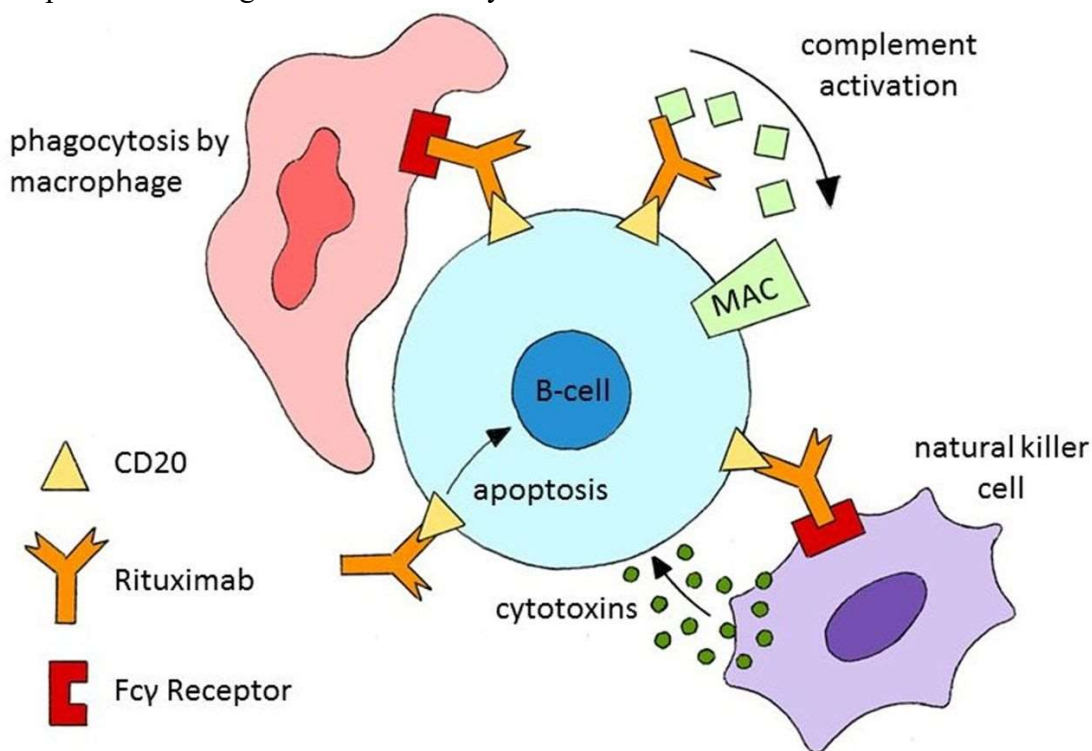
Generation	1 st Generation		2 nd Generation		3 rd Generation
mAb Structure 	Murine (100% rodent)  Suffix: - omab	Chimeric (65% human)  Suffix: - ximab	Humanised (>90% human)  Suffix: - zumab	Fully human  Suffix: - umab	Modified Fc region (chimeric or humanised) 
Immunogenicity	Higher  Lower				
Anti-CD20 mAbs	<i>Not in clinical use due to short half-life, poor efficacy and high risk of adverse reactions</i>	Rituximab Biosimilars: Truxima Rixathon <i>Unlicensed use in neurology (table 2)</i>	Ocrelizumab <i>Licensed for relapsing and primary progressive MS</i>	Ofatumumab <i>Currently in phase III clinical trials for MS</i>	Ublituximab (TG-1101) (chimeric) <i>Currently in phase III clinical trials for MS</i>
Anti-CD19 mAbs					Inebilizumab (MEDI-551) (humanised) <i>Currently in clinical trials for MS and NMOSD</i>

Figure 2: Neurology's use of B-cell-depleting monoclonal antibodies. Multiple sclerosis (MS), neuromyelitis optica spectrum disorder (NMOSD), monoclonal antibody.

The first anti-CD20 monoclonal antibody to be licensed for the treatment of B-cell lymphomas was rituximab (1997). Since then, it has been approved for the treatment of ANCA-associated vasculitis and refractory rheumatoid arthritis. The use of illegal drugs for neuroinflammatory diseases is expanding. Rituximab is a first-generation chimeric monoclonal antibody created by

combining a human Fc domain with a murine (rodent) Fab domain (the term "chimeric" comes from the mythical creature known as the Chimera, a terrible fire-breathing hybrid comprised of parts lion and goat). Figure 3 illustrates how the Fc domain stimulates numerous immunological responses.

Within three days of the first rituximab injection, 90% of the circulating B-cells are destroyed. In several illnesses, effectiveness is correlated with a decrease in pathogenic antibody titers. The full spectrum of B-cell function is likely to be affected by rituximab, though, and secondary T-cell function modifications, like the activation of immunoregulatory T cells, may be significant in some neuroinflammatory illnesses. It is envisaged that sparing CD20-negative long-lived plasma cells will protect enduring humoral immunity.



MAC membrane attack complex

Figure 3: Rituximab eliminates CD20+ B-cells by three separate pathways: (1) complement-dependent cytotoxicity; (2) antibody-dependent cellular cytotoxicity mediated by Fc receptors on the surface of natural killer cells, granulocytes, and macrophages; (3) activation of apoptosis.

Second-generation monoclonal antibodies feature enhanced Fab domains, which are frequently humanized or totally human and improve B-cell killing and tolerability, in comparison to first-generation monoclonal antibodies (figure 2). Recently, ocrelizumab (humanized) was authorized for the treatment of relapsing and progressive MS. Clinical trials are being conducted with ofatumumab, a completely human monoclonal antibody administered by monthly subcutaneous injection. The Fc-mediated immunological functions or half-life of third-generation monoclonal antibodies have been considerably enhanced. Currently being tested in MS is ubituximab (TG-1101), a fast infusible chimeric glycoengineered monoclonal antibody. Because CD19 is expressed

more widely throughout B-cell development, including the plasmablast phase, anti-CD19 B-cell-depleting treatments could be more effective (and possibly carry higher hazards) than anti-CD20 treatments (figure 1). A phase 3 trial of inebilizumab (MEDI-551) is being conducted in NMOSD.

Biosimilars

Monoclonal antibodies are typically expensive. However, once the patent on the original medication expires, less expensive copies, or "biosimilars," become accessible. Competing businesses are unable to access the original molecular clone, cell bank, or precise manufacturing procedure, which could cause these intricate molecular structures to change somewhat. Because of this, biosimilars are not really "generic." Biosimilars must be proved to be highly similar to the original monoclonal antibody in terms of structure, purity, and biological activity in order to obtain a license; however, it is not required to perform clinical trials for every indication. The European Medicines Agency (EMA) has approved two biosimilars, Truxima and Rixathon, since the patent for Rituximab expired in 2016. The administration and dosing procedures are the same. Currently, MabThera 1 g (the original version) costs £1746 according to the British National Formulary.

Rituximab indications and supporting data in neurology

Off-license prescribing should be guided by knowledge of the evidence for rituximab in neuroinflammatory diseases (see table 1 for a succinct summary).

Table 1. Indications for rituximab in neurology

INDICATION
1. Refractory Myasthenia Gravis
2. NMO-SD
3. Autoimmune encephalitis
4. Relapsing Remitting Multiple sclerosis
5. Immune-mediated Peripheral neuropathies
6. Primary angitis of the CNS
7. Stiff-person syndrome

Multiple sclerosis

Use of rituximab for MS in the UK is uncommon due to the availability of licensed disease-modifying treatments supported by phase III randomised controlled trials. But there is evidence that it works, so it might be a choice occasionally (especially if licensed comorbidities, such as active rheumatoid arthritis, facilitate funding). Rituximab's phase I and phase II trials for relapsing-remitting MS met their main objectives. 7–9 While a subgroup study of a major 96-week multicenter randomised controlled trial in primary progressive MS failed to reveal a delay to verified disease progression, younger individuals, especially those with inflammatory lesions, benefited from it. 10 Then, MS clinical trials came to a halt, most likely as a result of the approaching patent expiration of rituximab and the introduction of more recent B-cell depleting

medications from the same manufacturer.

In a large multicenter cohort (n=822), Sweden has published class IV evidence of safety and efficacy, making it the largest off-licence prescriber of rituximab for all forms of MS. 11 The dosage is 500–1000 mg six to twelve times a month. An efficacy in relapsing-remitting multiple sclerosis (MS) comparable to natalizumab and fingolimod, and significantly superior than injectable disease-modifying treatments and dimethyl fumarate, was demonstrated in a recent real-world retrospective comparison analysis. In terms of discontinuation rate, rituximab was superior to all other medications. 12 Despite the fact that this evidence is of rather low quality, there is a strong signal that rituximab is a successful treatment for MS, which is what one would anticipate given the recent encouraging randomised controlled trials for ocrelizumab.

We take 15 patients with MS and see relapse after and before rituximab administration

MS PATIENTS	RELAPSES BEFORE RITUXIMAB	RELAPSES AFTER RITUXIMAB	P VALUE
15	8 PER YEAR	1 PER YEAR	SIGNIFICANT LESS THAN 0.05

Neuromyelitis optica spectrum disorders

Despite the fact that three such trials are under underway, no immunosuppressive medication for NMOSD has been verified by a high-quality randomised controlled study. Numerous, primarily retrospective case reports totaling more than 400 patients that demonstrate persistent decreases in the annualized relapse rate support the use of rituximab. We will cover several dosing tactics in "dosing and monitoring" because there are many different ones in use. According to a recent meta-analysis, the relapse rate has decreased by 79% on average. 13 Rituximab presently has the greatest evidence of any immunotherapy used in NMOSD, however patients in the UK still receive it as second-line therapy because of its relatively expensive cost. It is offered to patients who have relapsed despite receiving appropriate treatment with mycophenolate mofetil or azathioprine in combination with low-dose prednisolone. 14

We take 5. patients of NMO and see the result after and before rituximab

NMO PATIENTS	RELAPSES BEFORE RITUXIMAB	RELAPSES AFTER RITUXIMAB	P VALUE
5	3PER YEAR	ZERO PER YEAR	SIGNIFICANT LESS THAN 0.05

Autoimmune encephalitis

Rituximab is typically used as a second-line acute therapy (single course) to maximize neurological recovery rather than as a long-term maintenance medication (as with MS/NMOSD)

because the majority of autoimmune encephalitis is monophasic. The most typical dosing schedule consists of four doses of 375 mg/m² given weekly. When there has been a poor response to intravenous corticosteroids, plasma exchange, and intravenous immunoglobulin, limited retrospective evidence supports its usage. It is impossible to attribute therapeutic advantages simply to rituximab because there is insufficient information to compare the impact of various immunotherapies in autoimmune encephalitis. However, it is an appealing choice due to its quick onset of action, proven efficacy in other antibody-mediated illnesses, and acceptable safety profile with short-term usage. The primary investigation in favor of rituximab usage in autoimmune encephalitis is a retrospective comparison of 161 patient outcomes for encephalitis. Regardless of antibody status, functional improvement as judged by the modified Rankin Scale happened more frequently in the rituximab-treated group. 15

Additional data are available specifically for the most prevalent subtype of autoimmune encephalitis, anti-NMDAR encephalitis. Compared to 55% of patients who failed first-line and did not receive second-line immunotherapy, a large prospective cohort study (n=577) indicated that 78% of patients who failed first-line and got second-line immunotherapy (rituximab and/or cyclophosphamide) had a satisfactory result at 24 months. 16 44 patients with anti-NMDAR encephalitis participated in a rituximab research in pediatric neuroinflammatory illness. Second-line rituximab therapy, especially when started early, helped 97% of these patients in some way. 17 Due to these results, a UK clinical commissioning strategy that was released in March 2018 decided to routinely fund rituximab for adults and kids with anti-NMDAR encephalitis who have responded poorly to first-line therapy (failure to improve by six months after starting treatment). two or more points on the modified Rankin Scale by 4 weeks after beginning first-line therapy (or two or more points on the modified Rankin Scale by six weeks after the onset of symptoms). 18 Case reports and tiny case series, which are usually complicated by the coadministration of various immunotherapies, are the only sources of evidence for autoimmune encephalitis caused by less prevalent antibodies. Two case series, for instance, detail the results of rituximab in seven patients with anti-LGI1 encephalitis. One patient may have responded, while three patients (43%) had successful outcomes. 19 20 Regardless of the presence of antibodies, early and intensive immunotherapy is useful in autoimmune encephalitis. It is likely that immunotherapy algorithms will increasingly include rituximab or other B-cell-depleting medications.

Primary angiitis of the central nervous system

The majority of treatment for this uncommon illness is high-dose corticosteroids, either in combination with or without cyclophosphamide. 21 Rituximab has produced positive results in two small case series, where 2/2 and 6/7 of the patients showed signs of improvement. 22 23 Additional case studies describing its application are available. 24

ANCA-associated vasculitis

Mononeuritis multiplex is one way that ANCA-associated vasculitis may infrequently present to the neurologist, but this condition is usually co-managed by other vasculitis specialists. Recent European Guidelines for organ or life-threatening disease license and recommend rituximab. 25 Rituximab (375 mg/m² weekly for four doses) was non-inferior to cyclophosphamide for

producing remission in two randomised controlled studies. 26 27 For relapsing disease, it might be more effective than cyclophosphamide. 27 Rituximab will be covered by NHS England where cyclophosphamide has failed or is not appropriate (eg, patients who wish to preserve their reproductive potential). 28

Stiff-person syndrome

A single small double-blind randomised controlled trial (n=24) reported no significant changes in any outcome measures following 6 months of rituximab treatment, despite some case reports suggesting a potential benefit for stiff-person syndrome. 32

Myasthenia gravis

Rituximab should be taken into consideration as an early therapeutic option in patients with MuSK-associated myasthenia gravis who have an inadequate response to first immunotherapy, suggest international consensus guidelines from 2016. 51 For myasthenia gravis related with the AChR, a formal consensus could not be obtained. Rituximab has been studied as an acute therapy (often a single course with varying doses) for refractory myasthenia gravis in a number of primarily retrospective, observational studies and two systematic reviews (persistent weakness or need for high-dose corticosteroids despite conventional immunosuppression).

The reported response rates in AChR-associated myasthenia gravis are inconsistent, with 30%–80% of patients achieving a Myasthenia Gravis Foundation of America post-intervention status (MGFA-PIS) of "minimal manifestations or better" after rituximab, despite many case series being shared between the systematic reviews. 52 53 The variation in patient selection, the inclusion of numerous "burned out," non-responsive instances, and the inclusion of cases where MGFA-PIS was not utilized as an end measure in the original publication may all help to explain this. Response and AChR antibody titres did not correspond well. 53 In the near future, two current randomised controlled trials may contribute to a better understanding of the effectiveness of rituximab in AChR-associated myasthenia gravis.

In contrast, both evaluations showed high (72%–89%) response rates in MuSK-associated myasthenia gravis. 52 53 A second blinded prospective assessment indicated that 26% of controls and 67% of rituximab-treated patients had MGFA-PIS of "limited symptoms or better." 54 In MuSK-associated myasthenia gravis, the benefit of rituximab seems to last longer and correlate better with antibody titres. 53 55 Unlike AChR antibodies, which are of the IgG1/3 subtype, MuSK antibodies are of the IgG4 subtype. Therefore, the selective eradication of transient IgG4-producing B-cells may account for rituximab's greater effectiveness. 55

We take 10 patients of MG and see the result after and before rituximab

MG PATIENTS	RELAPSES BEFORE RITUXIMAB	RELAPSES AFTER RITUXIMAB	P VALUE
10	4PER YEAR	ZERO PER YEAR	SIGNIFICANT LESS THAN 0.05

Treatment not working

When treatment failure is suspected, we advise ruling out other explanations, such as concurrent infection, and confirming that B-cell depletion is sufficient by looking at the CD19+ B-cell count in peripheral blood. Following are some potential causes of therapy failure:

1. Ineffectiveness of B-cell depletion

Nine patients (9%) in a large NMOSD cohort (n=100) relapsed after target-range CD19+/CD27+ memory B-cell depletion.

When taking rituximab, NMOSD relapses are typically less severe than when not taking it. It is not believed that rituximab depletes long-lived plasma cells or non-circulating B-cells in lymphoid organs, which together make up the majority of the body's total B-cell population and may have a role in breakthrough disease.

2. *Early relapses/delayed therapeutic onset*

Rituximab induction therapy may be followed by early NMOSD relapses. 4 72 73 This might be the result of insufficient B-cell depletion. Alternately, initial B-cell depletion may cause the release of systemic B-cell activation factor, encouraging plasma cells to produce autoantibodies, "resulting in a transitory spike" in antibody titre, and triggering early relapses. 74

3. *Incomplete B-cell depletion/early repopulators*

Rituximab induction therapy may be followed by early NMOSD relapses. 4 72 73 This might be the result of insufficient B-cell depletion. Alternately, initial B-cell depletion may cause the release of systemic B-cell activation factor, encouraging plasma cells to produce autoantibodies, "resulting in a transitory spike" in antibody titre, and triggering early relapses. 74

4. *Antidrug antibodies*

Some monoclonal antibodies, such as anti-tumor necrosis factor agents, have less efficacy because they are reactive to other drugs. Fc binding might improve medication clearance while Fab binding might have a neutralizing impact. Anti-drug antibodies may have a part in rituximab treatment failure, however this is unclear. They were discovered in one-third of the MS patients receiving rituximab treatment. 76 Antidrug antibodies may have a bigger impact in individuals receiving low-dose rituximab (100 mg infusions)⁷⁷, but higher, conventional dosages will likely mitigate their effects. 76 78 Antidrug antibodies can be technically challenging, poorly standardized, and difficult to produce for everyday use outside of clinical studies.

Combination with other immunosuppressive medications

Due to the risk of early relapse after rituximab initiation, some neurologists continue moderate-dose prednisolone (usually 10–20 mg daily) for 4–12 weeks in NMOSD. The decision to continue corticosteroids depends on the condition being treated and individual patient factors.

Combination with other immunosuppressive medications can be considered in some circumstances but must be balanced against the risk of immunocompromise. We generally reserve combination therapy for refractory disease. In treating rheumatoid arthritis, rituximab is often combined with

methotrexate or leflunomide but there is little evidence to guide practice in neuroinflammatory disease.

Pregnancy and breast feeding

The placenta is crossed by rituximab after 20 weeks of pregnancy. The information currently available shows that rituximab may be safe to use during early pregnancy, while this is not known for sure.⁸² It is possible to take advantage of the prolonged B-cell depleting impact, which can sometimes last longer than 40 weeks of gestation. Rituximab, for instance, might be administered both before and after delivery in pregnancies that were planned, protecting the developing fetus against B-cell depletion.

The danger of recurrence after prolonged cessation of rituximab medication for conception and pregnancy is a conundrum for many women with relapsing illnesses with substantial morbidity, such as NMOSD. In the hopes that B-cell depletion may occur, a new expert study proposes that two doses of 1000 mg could be administered as soon as one month before anticipated conception. The depletion will continue throughout the pregnancy. Given the extremely high postpartum risk of NMOSD relapse, they advise that rituximab could be begun during the first week of birth.⁸³ However, women should receive advice with the scant information on pregnancies exposed to rituximab.⁸⁴

Conclusion

The depletion will continue throughout the pregnancy. They recommend rituximab as an effective treatment for a number of neuroinflammatory disorders. There is rising evidence to support its usage in certain circumstances, even if there are no randomised controlled trials and doubts about the best dose methods. Overall, rituximab has a very good safety record, and compared to other immunomodulatory therapies, it may be a viable alternative for treating severe active illnesses during pregnancy. Neurologists, however, need to be mindful of certain management difficulties, such as secondary antibody deficit in patients needing maintenance B-cell depletion. Low pretreatment immunoglobulin levels, prior immunosuppressive medication use, or a need for continuous combination therapy are some specific risk factors to take into account. In the first week, ximab treatment might be restarted.

The efficacy and duration of more recent and ongoing B-cell depleting medications have shown significant promise, but it is yet unknown whether these will carry new dangers. The real-world hazards and benefits for patients will need to be more clearly defined, which will require prospective registries with long-term follow-up.

References

- 1) Perez-Andres M , Paiva B , Nieto WG , et al . Human peripheral blood B-cell compartments: a crossroad in B-cell traffic. *Cytometry B Clin Cytom* 2010;78(S1):47–60.doi:10.1002/cyto.b.20547 Google Scholar
- 2) Magliozzi R , Howell O , Vora A , et al . Meningeal B-cell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical

- pathology. *Brain* 2007;130(Pt 4):1089–104.doi:10.1093/brain/awm038 CrossRefPubMedWeb of ScienceGoogle Scholar
- 3) Baker D , Marta M , Pryce G , et al . Memory B-cells are major targets for effective immunotherapy in relapsing multiple sclerosis. *EBioMedicine* 2017;16:41–50.doi:10.1016/j.ebiom.2017.01.042 Google Scholar
 - 4) Kim SH , Kim W , Li XF , et al . Repeated treatment with rituximab based on the assessment of peripheral circulating memory B-cells in patients with relapsing neuromyelitis optica over 2 years. *Arch Neurol* 2011;68:1412–9.doi:10.1001/archneurol.2011.154 CrossRefPubMedWeb of ScienceGoogle Scholar
 - 5) Cree BA , Bennett JL , Sheehan M , et al . Placebo-controlled study in neuromyelitis optica-ethical and design considerations. *Mult Scler* 2016;22:862–72.doi:10.1177/1352458515620934 CrossRefPubMedGoogle Scholar
 - 6) British National Formulary: Rituximab, 2018. <https://bnf.nice.org.uk/drug/rituximab.html> [accessed 7th Mar].Google Scholar
 - 7) Bar-Or A , Calabresi PA , Arnold D , et al . Rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial. *Ann Neurol* 2008;63:395–400.doi:10.1002/ana.21363 CrossRefPubMedWeb of ScienceGoogle Scholar
 - 8) Hauser SL , Waubant E , Arnold DL , et al . B-cell depletion with rituximab in relapsing-remitting multiple sclerosis. *N Engl J Med Overseas Ed* 2008;358:676–88.doi:10.1056/NEJMoa0706383 Google Scholar
 - 9) Naismith RT , Piccio L , Lyons JA , et al . Rituximab add-on therapy for breakthrough relapsing multiple sclerosis: a 52-week phase II trial. *Neurology* 2010;74:1860–7.doi:10.1212/WNL.0b013e3181e24373 CrossRefPubMedGoogle Scholar
 - 10) Hawker K , O'Connor P , Freedman MS , et al . Rituximab in patients with primary progressive multiple sclerosis: results of a randomized double-blind placebo-controlled multicenter trial. *Ann Neurol* 2009;66:460–71.doi:10.1002/ana.21867 CrossRefPubMedWeb of ScienceGoogle Scholar
 - 11) Salzer J , Svenningsson R , Alping P , et al . Rituximab in multiple sclerosis: a retrospective observational study on safety and efficacy. *Neurology* 2016;87:2074–81.doi:10.1212/WNL.0000000000003331 Google Scholar
 - 12) Granqvist M , Borelmalm M , Poorghobad A , et al . Comparative effectiveness of rituximab and other initial treatment choices for multiple sclerosis. *JAMA Neurol* 2018;75:320–7.doi:10.1001/jamaneurol.2017.4011 Google Scholar
 - 13) Damato V , Evoli A , Iorio R . Efficacy and safety of rituximab therapy in neuromyelitis optica spectrum disorders: a systematic review and meta-analysis. *JAMA Neurol* 2016;73:1342–8.doi:10.1001/jamaneurol.2016.1637 Google Scholar
 - 14) Palace J , Leite MI , Leite I , et al . A practical guide to the treatment of neuromyelitis optica. *Pract Neurol* 2012;12:209–14.doi:10.1136/practneurol-2012-000237 FREE Full TextGoogle Scholar

- 15) Lee WJ , Lee ST , Byun JI , et al . Rituximab treatment for autoimmune limbic encephalitis in an institutional cohort. *Neurology* 2016;86:1683–91.doi:10.1212/WNL.0000000000002635 Google Scholar
- 16) Titulaer MJ , McCracken L , Gabilondo I , et al . Treatment and prognostic factors for long-term outcome in patients with anti-NMDA receptor encephalitis: an observational cohort study. *Lancet Neurol* 2013;12:157–65.doi:10.1016/S1474-4422(12)70310-1 CrossRefPubMedWeb of ScienceGoogle Scholar
- 17) Dale RC , Brilot F , Duffy LV , et al . Utility and safety of rituximab in pediatric autoimmune and inflammatory CNS disease. *Neurology* 2014;83:142–50.doi:10.1212/WNL.0000000000000570 CrossRefPubMedGoogle Scholar
- 18) Clinical Commissioning Policy, 2018. Rituximab for second line treatment for anti-NMDAR autoimmune encephalitis (all ages). NHS England. <https://www.england.nhs.uk/wp-content/uploads/2018/03/ccp-rituximab-for-second-line-treatment-for-anti-nmdar-autoimmune-encephalitis.pdf> Google Scholar
- 19) Brown JW , Martin PJ , Thorpe JW , et al . Long-term remission with rituximab in refractory leucine-rich glioma inactivated 1 antibody encephalitis. *J Neuroimmunol* 2014;271:66–8.doi:10.1016/j.jneuroim.2014.03.012 PubMedGoogle Scholar
- 20) Irani SR , Gelfand JM , Bettcher BM , et al . Effect of rituximab in patients with leucine-rich, glioma-inactivated 1 antibody-associated encephalopathy. *JAMA Neurol* 2014;71:896–900.doi:10.1001/jamaneurol.2014.463 Google Scholar
- 21) Salvarani C , Brown RD , Christianson TJ , et al . Adult primary central nervous system vasculitis treatment and course: analysis of one hundred sixty-three patients. *Arthritis Rheumatol* 2015;67:1637–45.doi:10.1002/art.39068 Google Scholar
- 22) ¢Berlit P , Becker J , Kraemer M . Rituximab in primary angiitis of the CNS (P3.076). *Neurology* 2017;88.Google Scholar
- 23) ¢De Boysson H , Arquizan C , Guillevin L , et al . Rituximab for primary angiitis of the central nervous system: report of 2 patients from the French COVAC cohort and review of the literature. *J Rheumatol* 2013;40:2102–3.doi:10.3899/jrheum.130529 FREE Full TextGoogle Scholar
- 24) ¢Salvarani C , Brown RD , Huston J , et al . Treatment of primary CNS vasculitis with rituximab: case report. *Neurology* 2014;82:1287–8.doi:10.1212/WNL.0000000000000293 Google Scholar
- 25) ¢Yates M , Watts RA , Bajema IM , et al . EULAR/ERA-EDTA recommendations for the management of ANCA-associated vasculitis. *Ann Rheum Dis* 2016;75:1583–94.doi:10.1136/annrheumdis-2016-209133 Abstract/FREE Full TextGoogle Scholar
- 26) ¢Jones RB , Furuta S , Tervaert JW , et al . Rituximab versus cyclophosphamide in ANCA-associated renal vasculitis. *Ann Rheum Dis* 2015;74:1178–82.Abstract/FREE Full TextGoogle Scholar

- 27) Stone JH, Merkel PA, Spiera R, et al. Rituximab versus cyclophosphamide for ANCA-associated vasculitis. *N Engl J Med* 2010;363:221–32.doi:10.1056/NEJMoa0909905 CrossRefPubMedWeb of ScienceGoogle Scholar
- 28) National Institute for Health and Care Excellence technology appraisal guidance TA308, 2014. Rituximab in combination with glucocorticoids for treating anti-neutrophil cytoplasmic antibody-associated vasculitis. <https://www.nice.org.uk/guidance/ta308/chapter/1-Guidance> [Accessed 16 Jun 2018].Google Scholar
- 29) Baker MR, Das M, Isaacs J, et al. Treatment of stiff person syndrome with rituximab. *J Neurol Neurosurg Psychiatry* 2005;76:999–1001.doi:10.1136/jnnp.2004.051144 Abstract/FREE Full TextGoogle Scholar
- 30) Dupond JL, Essalmi L, Gil H, et al. Rituximab treatment of stiff-person syndrome in a patient with thymoma, diabetes mellitus and autoimmune thyroiditis. *J Clin Neurosci* 2010;17:389–91.doi:10.1016/j.jocn.2009.06.015 CrossRefPubMedGoogle Scholar
- 31) Katoh N, Matsuda M, Ishii W, et al. Successful treatment with rituximab in a patient with stiff-person syndrome complicated by dysthyroid ophthalmopathy. *Intern Med* 2010;49:237–41.doi:10.2169/internalmedicine.49.2821 PubMedGoogle Scholar
- 32) Dalakas MC, Rakocevic G, Dambrosia JM, et al. A double-blind, placebo-controlled study of rituximab in patients with stiff person syndrome. *Ann Neurol* 2017;82:271–7.doi:10.1002/ana.25002 Google Scholar
- 33) Clinical Commissioning Policy, 2017. Rituximab for chronic inflammatory demyelinating polyradiculoneuropathy (CIDP), multifocal motor neuropathy (MMN), vasculitis of the peripheral nervous system & IgM paraprotein- associated demyelinating neuropathy (adults). NHS England. <https://www.england.nhs.uk/publication/commissioning-policy-rituximab-for-chronic-inflammatory-demyelinating-polyradiculoneuropathy-cidp-multifocal-motor-neuropathy-mm-n-vasculitis-of-the-peripheral-nervous-system-igm-paraprotein-a/> Google Scholar
- 34) Mahdi-Rogers M, van Doorn PA, Hughes RA. Immunomodulatory treatment other than corticosteroids, immunoglobulin and plasma exchange for chronic inflammatory demyelinating polyradiculoneuropathy. *Cochrane Database Syst Rev* 2013;6:CD003280.doi:10.1002/14651858.CD003280.pub4 Google Scholar
- 35) Benedetti L, Briani C, Franciotta D, et al. Rituximab in patients with chronic inflammatory demyelinating polyradiculoneuropathy: a report of 13 cases and review of the literature. *J Neurol Neurosurg Psychiatry* 2011;82:306–8.doi:10.1136/jnnp.2009.188912 Abstract/FREE Full TextGoogle Scholar
- 36) Cocito D, Grimaldi S, Paolasso I, et al. Immunosuppressive treatment in refractory chronic inflammatory demyelinating polyradiculoneuropathy. A nationwide retrospective analysis. *Eur J Neurol* 2011;18:1417–21.doi:10.1111/j.1468-1331.2011.03495.x CrossRefPubMedGoogle Scholar

- 37) Querol L , Rojas-García R , Diaz-Manera J , et al . Rituximab in treatment-resistant CIDP with antibodies against paranodal proteins. *Neurol Neuroimmunol Neuroinflamm* 2015;2:149.doi:10.1212/NXI.0000000000000149 Google Scholar
- 38) Delmont E , Manso C , Querol L , et al . Autoantibodies to nodal isoforms of neurofascin in chronic inflammatory demyelinating polyneuropathy. *Brain* 2017;140:1851–8.doi:10.1093/brain/awx124 Google Scholar
- 39) Levine TD , Pestronk A . IgM antibody-related polyneuropathies: b-cell depletion chemotherapy using Rituximab. *Neurology* 1999;52:1701–4.doi:10.1212/WNL.52.8.1701 CrossRefPubMedGoogle Scholar
- 40) Stieglbauer K , Topakian R , Hinterberger G , et al . Beneficial effect of rituximab monotherapy in multifocal motor neuropathy. *Neuromuscul Disord* 2009;19:473–5.doi:10.1016/j.nmd.2009.04.013 PubMedGoogle Scholar
- 41) Chaudhry V , Cornblath DR . An open-label trial of rituximab (Rituxan®) in multifocal motor neuropathy. *J Peripher Nerv Syst* 2010;15:196–201.doi:10.1111/j.1529-8027.2010.00270.x PubMedGoogle Scholar
- 42) Gorson KC , Natarajan N , Ropper AH , et al . Rituximab treatment in patients with IVIg-dependent immune polyneuropathy: a prospective pilot trial. *Muscle Nerve* 2007;35:66–9.doi:10.1002/mus.20664 CrossRefPubMedWeb of ScienceGoogle Scholar
- 43) Collins MP , Dyck PJ , Gronseth GS , et al . Peripheral nerve society guideline on the classification, diagnosis, investigation, and immunosuppressive therapy of non-systemic vasculitic neuropathy: executive summary. *J Peripher Nerv Syst* 2010;15:176–84.doi:10.1111/j.1529-8027.2010.00281.x CrossRefPubMedGoogle Scholar
- 44) Blaes F . Diagnosis and therapeutic options for peripheral vasculitic neuropathy. *Ther Adv Musculoskelet Dis* 2015;7:45–55.doi:10.1177/1759720X14566617 CrossRefPubMedGoogle Scholar
- 45) Dalakas MC , Rakocevic G , Salajegheh M , et al . Placebo-controlled trial of rituximab in IgM anti-myelin-associated glycoprotein antibody demyelinating neuropathy. *Ann Neurol* 2009;65:286–93.doi:10.1002/ana.21577 CrossRefPubMedWeb of ScienceGoogle Scholar
- 46) Léger JM , Viala K , Nicolas G , et al . Placebo-controlled trial of rituximab in IgM anti-myelin-associated glycoprotein neuropathy. *Neurology* 2013;80:2217–25.doi:10.1212/WNL.0b013e318296e92b CrossRefPubMedGoogle Scholar
- 47) Renaud S , Gregor M , Fuhr P , et al . Rituximab in the treatment of polyneuropathy associated with anti-MAG antibodies. *Muscle Nerve* 2003;27:611–5.doi:10.1002/mus.10359 CrossRefPubMedWeb of ScienceGoogle Scholar
- 48) Benedetti L , Briani C , Grandis M , et al . Predictors of response to rituximab in patients with neuropathy and anti-myelin associated glycoprotein immunoglobulin M. *J Peripher Nerv Syst* 2007;12:102–7.doi:10.1111/j.1529-8027.2007.00129.x CrossRefPubMedWeb of ScienceGoogle Scholar

- 49) «Benedetti L , Briani C , Franciotta D , et al . Long-term effect of rituximab in anti-mag polyneuropathy. *Neurology* 2008;71:1742–4.doi:10.1212/01.wnl.0000335268.70325.33 Google Scholar
- 50) «Niermeijer JM , Eurelings M , Lokhorst HL , et al . Rituximab for polyneuropathy with IgM monoclonal gammopathy. *J Neurol Neurosurg Psychiatry* 2009;80:1036–9.doi:10.1136/jnnp.2008.155325 Abstract/FREE Full TextGoogle Scholar
- 51) «Sanders DB , Wolfe GI , Benatar M , et al . International consensus guidance for management of myasthenia gravis: Executive summary. *Neurology* 2016;87:419–25.doi:10.1212/WNL.0000000000002790 Google Scholar
- 52) «Iorio R , Damato V , Alboini PE , et al . Efficacy and safety of rituximab for myasthenia gravis: a systematic review and meta-analysis. *J Neurol* 2015;262:1115–9.doi:10.1007/s00415-014-7532-3 PubMedGoogle Scholar
- 53) «Tandan R , Hehir MK , Waheed W , et al . Rituximab treatment of myasthenia gravis: a systematic review. *Muscle Nerve* 2017;56:185–96.doi:10.1002/mus.25597 Google Scholar
- 54) «Hehir MK , Hobson-Webb LD , Benatar M , et al . Rituximab as treatment for anti-MuSK myasthenia gravis: multicenter blinded prospective review. *Neurology* 2017;89:1069–77.doi:10.1212/WNL.0000000000004341 Google Scholar
- 55) «Díaz-Manera J , Martínez-Hernández E , Querol L , et al . Long-lasting treatment effect of rituximab in MuSK myasthenia. *Neurology* 2012;78:189–93.doi:10.1212/WNL.0b013e3182407982 CrossRefPubMedGoogle Scholar
- 56) «Mabthera SmPC, 2018. <https://www.medicines.org.uk/emc/product/3801/smpc> [Accessed 7 Mar 2018].Google Scholar
- 57) «Bredemeier M , de Oliveira FK , Rocha CM . Low- versus high-dose rituximab for rheumatoid arthritis: a systematic review and meta-analysis. *Arthritis Care Res* 2014;66:228–35.doi:10.1002/acr.22116 CrossRefWeb of ScienceGoogle Scholar
- 58) «Vital EM , Rawstron AC , Dass S , et al . Reduced-dose rituximab in rheumatoid arthritis: efficacy depends on degree of B-cell depletion. *Arthritis Rheum* 2011;63:603–8.doi:10.1002/art.30152 CrossRefPubMedWeb of ScienceGoogle Scholar
- 59) «Nielsen AS , Miravalle A , Langer-Gould A , et al . Maximally tolerated versus minimally effective dose: the case of rituximab in multiple sclerosis. *Mult Scler* 2012;18:377–8.doi:10.1177/1352458511418631 CrossRefPubMedGoogle Scholar
- 60) «Yang CS , Yang L , Li T , et al . Responsiveness to reduced dosage of rituximab in chinese patients with neuromyelitis optica. *Neurology* 2013;81:710–3.doi:10.1212/WNL.0b013e3182a1aac7 CrossRefPubMedGoogle Scholar
- 61) «Zhang M , Zhang C , Bai P , et al . Effectiveness of low dose of rituximab compared with azathioprine in chinese patients with neuromyelitis optica: an over 2-year follow-up study. *Acta Neurol Belg* 2017;117:695–702.doi:10.1007/s13760-017-0795-6 Google Scholar

- 62) Wang BJ , Wang CJ , Zeng ZL , et al . Lower dosages of rituximab used successfully in the treatment of anti-NMDA receptor encephalitis without tumour. *J Neurol Sci* 2017;377:127–32.doi:10.1016/j.jns.2017.04.007 Google Scholar
- 63) Greenberg BM , Graves D , Remington G , et al . Rituximab dosing and monitoring strategies in neuromyelitis optica patients: creating strategies for therapeutic success. *Mult Scler* 2012;18:1022–6.doi:10.1177/1352458511432896 CrossRefPubMedGoogle Scholar
- 64) Yi JS , Decroos EC , Sanders DB , et al . Prolonged B-cell depletion in MuSK myasthenia gravis following rituximab treatment. *Muscle Nerve* 2013;48:992–3.doi:10.1002/mus.24063 PubMedGoogle Scholar
- 65) Pellkofer HL , Krumbholz M , Berthele A , et al . Long-term follow-up of patients with neuromyelitis optica after repeated therapy with rituximab. *Neurology* 2011;76:1310–5.doi:10.1212/WNL.0b013e3182152881 CrossRefPubMedGoogle Scholar
- 66) Mealy MA , Wingerchuk DM , Palace J , et al . Comparison of relapse and treatment failure rates among patients with neuromyelitis optica: multicenter study of treatment efficacy. *JAMA Neurol* 2014;71:324–30.doi:10.1001/jamaneurol.2013.5699 Google Scholar
- 67) Kim SH , Huh SY , Lee SJ , et al . A 5-year follow-up of rituximab treatment in patients with neuromyelitis optica spectrum disorder. *JAMA Neurol* 2013;70:1110–7.doi:10.1001/jamaneurol.2013.3071 Google Scholar
- 68) Cohen M , Romero G , Bas J , et al . Monitoring CD27+ memory B-cells in neuromyelitis optica spectrum disorders patients treated with rituximab: Results from a bicentric study. *J Neurol Sci* 2017;373:335–8.doi:10.1016/j.jns.2017.01.025 Google Scholar
- 69) Collongues N , de Seze J . An update on the evidence for the efficacy and safety of rituximab in the management of neuromyelitis optica. *Ther Adv Neurol Disord* 2016;9:180–8.doi:10.1177/1756285616632653 CrossRefGoogle Scholar
- 70) Pittock SJ , Lucchinetti CF . Neuromyelitis optica and the evolving spectrum of autoimmune aquaporin-4 channelopathies: a decade later. *Ann N Y Acad Sci* 2016;1366:20–39.doi:10.1111/nyas.12794 Google Scholar
- 71) Kim SH , Jeong IH , Hyun JW , et al . Treatment outcomes with rituximab in 100 patients with neuromyelitis optica: Influence of FCGR3A polymorphisms on the therapeutic response to rituximab. *JAMA Neurol* 2015;72:989–95.doi:10.1001/jamaneurol.2015.1276 Google Scholar
- 72) Lindsey JW , Meulmester KM , Brod SA , et al . Variable results after rituximab in neuromyelitis optica. *J Neurol Sci* 2012;317:103–5.doi:10.1016/j.jns.2012.02.017 CrossRefPubMedGoogle Scholar
- 73) Perumal JS , Kister I , Howard J , et al . Disease exacerbation after rituximab induction in neuromyelitis optica. *Neurology - Neuroimmunology Neuroinflammation* 2015;2:61.doi:10.1212/NXI.0000000000000061 Google Scholar

- 74) Nakashima I , Takahashi T , Cree BA , et al . Transient increases in anti-aquaporin-4 antibody titers following rituximab treatment in neuromyelitis optica, in association with elevated serum BAFF levels. *J Clin Neurosci* 2011;18:997–8.doi:10.1016/j.jocn.2010.12.011 CrossRefPubMedGoogle Scholar
- 75) Ruysen-Witrand A , Rouanet S , Combe B , et al . Association between -871C>T promoter polymorphism in the B-cell activating factor gene and the response to rituximab in rheumatoid arthritis patients. *Rheumatology* 2013;52:636–41.doi:10.1093/rheumatology/kes344 CrossRefPubMedGoogle Scholar
- 76) Dunn N , Juto A , Ryner M , et al . Rituximab in multiple sclerosis: Frequency and clinical relevance of anti-drug antibodies. *Mult Scler* 2018;24:1224–33.doi:10.1177/1352458517720044 Google Scholar
- 77) Li T , Zhang LJ , Zhang QX , et al . Anti-rituximab antibody in patients with NMOSDs treated with low dose rituximab. *J Neuroimmunol* 2018;316:107–11.doi:10.1016/j.jneuroim.2017.12.021 Google Scholar
- 78) van Vollenhoven RF , Emery P , Bingham CO , et al . Long-term safety of rituximab in rheumatoid arthritis: 9.5-year follow-up of the global clinical trial programme with a focus on adverse events of interest in RA patients. *Ann Rheum Dis* 2013;72:1496–502.doi:10.1136/annrheumdis-2012-201956 Abstract/FREE Full TextGoogle Scholar
- 79) De La Torre I , Leandro MJ , Valor L , et al . Total serum immunoglobulin levels in patients with RA after multiple B-cell depletion cycles based on rituximab: relationship with B-cell kinetics. *Rheumatology* 2012;51:833–40.doi:10.1093/rheumatology/ker417 CrossRefPubMedWeb of ScienceGoogle Scholar
- 80) Gottenberg JE , Ravaud P , Bardin T , et al . Risk factors for severe infections in patients with rheumatoid arthritis treated with rituximab in the autoimmunity and rituximab registry. *Arthritis Rheum* 2010;62:2625–32.doi:10.1002/art.27555 CrossRefPubMedWeb of ScienceGoogle Scholar
- 81) Tallantyre EC , Whittam DH , Paling D , et al . Secondary antibody deficiency and infection following B-cell depletion for CNS neuroinflammation. *J Neurol* 2018;265:1115–22.Google Scholar
- 82) Chakravarty EF , Murray ER , Kelman A , et al . Pregnancy outcomes after maternal exposure to rituximab. *Blood* 2011;117:1499–506.doi:10.1182/blood-2010-07-295444 Abstract/FREE Full TextGoogle Scholar
- 83) Shosha E , Pittock SJ , Flanagan E , et al . Neuromyelitis optica spectrum disorders and pregnancy: interactions and management. *Mult Scler* 2017;23:1808–17.doi:10.1177/1352458517740215 Google Scholar
- 84) Das G , Damotte V , Gelfand JM , et al . Rituximab before and during pregnancy: a systematic review, and a case series in MS and NMOSD. *Neurol Neuroimmunol Neuroinflamm* 2018;5:e453.doi:10.1212/NXI.0000000000000453 Google Scholar

- 85) «Hauser SL , Bar-Or A , Comi G , et al . Ocrelizumab versus interferon beta-1a in relapsing multiple sclerosis. N Engl J Med Overseas Ed 2017;376:221–34.doi:10.1056/NEJMoa1601277 Google Scholar
- 86) «Montalban X , Hauser SL , Kappos L , et al . Ocrelizumab versus placebo in primary progressive multiple sclerosis. N Engl J Med 2017;376:209–20.doi:10.1056/NEJMoa1606468 CrossRefPubMedGoogle Scholar
- 87) «Emery P , Rigby W , Tak PP . Safety with ocrelizumab in rheumatoid arthritis: results from the ocrelizumab phase iii pty with Ocrelizumab in Rheumatoid Arthritis: Results from the Ocrelizumab Phase III Program. PLoS One 2014;9:87379.doi:10.1371/journal.pone.0087379 Google Scholar
- 88) «Venhoff N , Effelsberg NM , Salzer U , et al . Impact of rituximab on immunoglobulin concentrations and B-cell numbers after cyclophosphamide treatment in patients with ANCA-associated vasculitides. PLoS One 2012;7:37626.doi:10.1371/journal.pone.0037626 Google Scholar
- 89) «Jolles S , Chapel H , Litzman J . When to initiate immunoglobulin replacement therapy (IGRT) in antibody deficiency: a practical approach. Clin Exp Immunol 2017;188:333–41.Google Scholar
- 90) «Chua I , Lagos M , Charalambous BM , et al . Pathogen-specific IgG antibody levels in immunodeficient patients receiving immunoglobulin replacement do not provide additional benefit to therapeutic management over total serum IgG. J Allergy Clin Immunol 2011;127:1410–1.doi:10.1016/j.jaci.2011.01.035 CrossRefPubMedWeb of ScienceGoogle Scholar
- 91) «Wimperis J , Lunn M , Jones A , 2011. Department of health clinical guidelines for immunoglobulin use: second edition update. Available from: https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/216671/dh_131107.pdf [Accessed Jul 2011].Google Scholar
- 92) Witzel SJ . Lactation and the use of biologic immunosuppressive medications. Breastfeed Med 2014;9:543–6.doi:10.1089/bfm.2014.0107 Google Scholar
- 93) Medicines and Healthcare products Regulatory Agency, 2013. Rituximab: screen for hepatitis B virus before treatment. MRHA Alert. Available from: <https://www.gov.uk/drug-safety-update/rituximab-screen-for-hepatitis-b-virus-before-treatment> Google Scholar
- 94) Hwang JP , Lok AS . Management of patients with hepatitis B who require immunosuppressive therapy. Nat Rev Gastroenterol Hepatol 2014;11:209–19.doi:10.1038/nrgastro.2013.216 PubMedGoogle Scholar
- 95) Sagnelli E , Pisaturo M , Sagnelli C , et al . Rituximab-based treatment, HCV replication, and hepatic flares. Clin Dev Immunol 2012;2012:1–5.doi:10.1155/2012/945950 CrossRefGoogle Scholar

- 96) Lin KM , Lin JC , Tseng WY , et al . Rituximab-induced hepatitis C virus reactivation in rheumatoid arthritis. *J Microbiol Immunol Infect* 2013;46:65–7.doi:10.1016/j.jmii.2011.12.020 CrossRefPubMedGoogle Scholar
- 97) Cantini F , Nannini C , Niccoli L , et al . Risk of tuberculosis reactivation in patients with rheumatoid arthritis, ankylosing spondylitis, and psoriatic arthritis receiving non-anti-TNF-targeted biologics. *Mediators Inflamm* 2017;2017:1–15.doi:10.1155/2017/8909834 CrossRefGoogle Scholar
- 98) Buch MH , Smolen JS , Betteridge N , et al . Updated consensus statement on the use of rituximab in patients with rheumatoid arthritis. *Ann Rheum Dis* 2011;70:909–20.doi:10.1136/ard.2010.144998 Abstract/FREE Full TextGoogle Scholar
- 99) Bingham CO , Looney RJ , Deodhar A , et al . Immunization responses in rheumatoid arthritis patients treated with rituximab: results from a controlled clinical trial. *Arthritis Rheum* 2010;62:64–74.doi:10.1002/art.25034 CrossRefPubMedWeb of ScienceGoogle Scholar
- 100) Kim W , Kim SH , Huh SY , et al . Reduced antibody formation after influenza vaccination in patients with neuromyelitis optica spectrum disorder treated with rituximab. *Eur J Neurol* 2013;20:975–80.doi:10.1111/ene.12132 CrossRefPubMedGoogle Scholar
- 101) Salmon JH , Cacoub P , Combe B , et al . Late-onset neutropenia after treatment with rituximab for rheumatoid arthritis and other autoimmune diseases: data from the AutoImmunity and Rituximab registry. *RMD Open* 2015;1:e000034.doi:10.1136/rmdopen-2014-000034 Abstract/FREE Full TextGoogle Scholar
- 102) Seibert J , Blackburn J , Vollmer B . Safety, tolerability, and efficacy of rituximab in the treatment of multiple sclerosis: 285 patients treated in a single center. *Neurology* 2015;84.Google Scholar
- 103) Plate A , Havla J , Kämpfel T . Late-onset neutropenia during long-term rituximab therapy in neuromyelitis optica. *Mult Scler Relat Disord* 2014;3:269–72.doi:10.1016/j.msard.2013.08.001 Google Scholar
- 104) Mealy MA , Levy M . Favorable outcome of granulocyte colony-stimulating factor use in neuromyelitis optica patients presenting with agranulocytosis in the setting of rituximab. *J Neuroimmunol* 2015;287:29–30.doi:10.1016/j.jneuroim.2015.08.003 Google Scholar
- 105) Reitblat T , Wechsler A , Reitblat O . Rituximab-related late-onset neutropenia in patients with rheumatic diseases: successful re-challenge of the treatment. *Am J Case Rep* 2015;16:211–4.doi:10.12659/AJCR.892541 Google Scholar
- 106) Zaheer F , Berger JR . Treatment-related progressive multifocal leukoencephalopathy: current understanding and future steps. *Ther Adv Drug Saf* 2012;3:227–39.doi:10.1177/2042098612453849 CrossRefPubMedGoogle Scholar